Instructions: Bold fields must be completed.

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End Time	Start Time		Collector(s)	Conductivity (ZM≥99 umhos/cm)	Scchi m)	AIS sign?	Date(s)	THE PROPERTY OF	Cour	WBIC	ame	Location N

STEP 1: Circle species that you looked for and review the Identification Handout.

}	AQUATIC PLANTS/ALGAE Hydrilla European frogbit Curly leaf Yellow floating heart Fanwort Brazilian waterweed Parrot fea
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sample of any new AIS found. Collect five new invasive plant specimens, 20 Dreissenids, and up to 3 of each invertebrate species. Include internal and external labels with STEP 2: Record locations of sampling sites (in decimal degrees). Indicate whether snorkeled or why not. List AIS found and density at each site or record none. Collect a WBIC, name of lake, county, sample date, sample type (snails, spiny water flea or zebra mussel) and collector. Legibility is appreciated. If needed, preserve with adequate

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	J.C. V. P. P. C.	************		-		a=12=	Alge	Snorkel If no, indicate (Y/N) why†
					Cons-1 live		const the	Species name, density (1-5)*, and live (L) or dead (D)
					~	ľ	W Y	Sample (Y/N)
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		\times	X	>		\times		No AIS
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^{*}boat landing (BL), target site (TS), meander survey (MS).

[†]Stained water, turbid water, blue-green bloom, chemical treatment, other (please describe).

⁹Live (L) animals will contain flesh and live plants will generally be rooted. Dead (D) animals will not contain flesh and dead plants include sterile fragments. *Density ratings: 1-a few plants or invertebrates, 2-one or a few plant beds or colonies of invertebrates, 3-many small beds or scattered plants or colonies of invertebrates, 4-dense plant, snail, or mussel growth in a while bay or portion of the lake, or 5-dense plant, snail or mussel growth covering most shallow areas.

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Harly Detection Monitoring Data Form

STEP 3: Collect Waterflea Tows from the deep hole (DH). Decant water and preserve the sample. Preserve with 4 parts ethanol and 1 part sample. Submit the sample, a completed copy of this data form, and a completed copy of the Water Flea Tow Monitoring Report (3200-128) to DNR Science Services. Legibility is appreciated.

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STEP 4: Collect vertical Veliger Tows from 3 sites; the deep hole (DH) and two other deep areas along the downwind side of the lake. Preserve with 4 parts ethanol and 1 part sample. Submit the sample, a copy of this completed data form, and a completed copy of the Mussel Veliger Tow Monitoring Report (3200-135) to DNR Science Service. Legibility is appreciated.

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^{*}Horizontal, oblique, or vertical

t30 or 50 cm.

‡Non-denatured or denatured ethanol

STEP 5: Coordinate voucher and sample submission and verification with regional DNR staff for all AIS records for the specific region.

- Freckmann Herbarium, Wisconsin State Herbarium, Other Plants will be compiled and entered into a spreadsheet to be verified and submitted to a herbarium by an in-person appointment. Please indicate which herbarium: _. Date of herbarium meeting
- Snails will be compiled with other regional snail specimens and sent to UW La Crosse. Date sent 1-25-2015
- Dreissenids will be sent to Science Services. Date sent

• Crayfish compiled and sent to: Craig Roesler or Scott VanEgeren. Date STEP 6: Data was entered into SWIMS on 7-28-2015

Once data is entered, send scans of data sheets to central office (Maureen.Ferry@Wisconsin.gov) and Amanda.Perdzock@Wisconsin.gov)

STEP 7: Data was proofed on