8/3/2/Y

Form 3200-xxx (R 6/2013)

				(-)		(40 0 / 2015)
Lake Name	County	WBIC	Date(s)	AIC class 2	1 600	
WILSON			Date(s) 7/22	AIS sign?	Secchi (ft or m)	Conductivity (ZM tow if > 99 umhos/cm)
1410013	MAUSHARA	75000	166	(Y) N	any from	
Data collectors		Load Monitor phase			suf I word	
AMY KRETU	WC	Lead Monitor phone and	d email Start time (~ 15 min)	End time (~ 15 min)	Total collector time (hrs x # collectors)
	×	950 BSB 637	9:15			" dille (1113 X # CONECCOIS)
CIBBY HESS		Elizabeth. Hoss @	1,1	1	1130 AM	1.25 hrs x 2 - z
L		MECONDINISOU	_	1		
LOOK for the following	snacias: Durnin lancast	rifo Dhu				

Look for the following species: Purple loosestrife, Phragmites, flowering rush, Japanese knotweed, Yellow iris, Eurasian water-milfoil, curly-leaf pondweed, Hydrilla, Brazillan waterweed, yellow floating heart, European frog-bit, yellow floating heart, water chestnut, Brazilian waterweed, fanwort, parrot feather, water hyacinth, water lettuce, zebra mussel, quagga mussel, water flea, Chinese mystery snail, banded mystery snail, faucet snail, New Zealand mud snail, Asian clam, red swamp crayfish, rusty crayfish, didymo, and any other AIS found.

STEP 1: Record locations of sampling sites (in decimal degrees). Sampling sites include all public boat landings (BL), 5 target sites (TS) and the meander survey sites (MS). List AIS found at each site or record none. Collect a sample of any new AIS found. Collect five new invasive plant specimens, 20 Dreissenids, and 3 of each snail species and include internal and external labels with WBIC, lake name, county, sample date, sample type (snails, spiny water flea or zebra mussel) and collector. Legibility is appreciated. If needed, preserve with adequate ethanol.

Site	Latitude	Longitude	Snorkel (Y or N*)	If N snorkel, indicate why	Species, density 1-5 [‡]
				i i i i i i i i i i i i i i i i i i i	DEAD / ALIVE
	44.17418	89.17270	4		P12-1-244-155 K
	4417231	-89.17424	4		ENM-2 SAMQUE BMS-2-SAMPCE LNM-2
1ch	44.17240	-89,7475	fracion)		CLP-1 SAMPLE
m52	44,7530	-89.17778	proved		PL -1
3	44.17-3660	39,17774	A.		BM5-2 EWM-1
M53	44,19678	491.7382	(m)		CZ 3
BIL	44. 17832	-41716	N	too hart fifty	DL-2, 80% 1, EV-11-2
Щ	44,17756	-99,17036	4	BEARY	CLEI ENM! BMS 2
5	44.17474	-89, 17010		David Pr	
(4)X	44.1726121	-87.1716034	N	Namon lovneh	EUR-1 EMW-1 ST-1

*For lakes	/sites no	t snorkele	d, substitute:
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Boat landing site – Examine rake throws and D-net samples for 30 minutes. Targeted site – Examine rake throws and D-net samples for 10 minutes. Meander – Examine 50 rake throws/D-net samples during meander survey.

†If lake/site was not snorkeled, indicate why: stained water, turbid water, blue-green bloom, chemical treatment, other (please describe).

‡ Density Ratings

1 – A few plants or invertebrates

4 – Dense plant, snail or mussel growth in a whole bay or portion of the lake

2 – One or a few plant beds or colonies of invertebrates

5 – Dense plant, snail or mussel growth covering most shallow areas

3 – Many small beds or scattered plants or colonies of invertebrates

Step 2: Collect Waterflea Tows from the deep hole (DH). Decant's water and preserve the sample. Submit the sample, this data form and the Water Flea Tow Monitoring Report (3200-128) to DNR Science Services.

Site	Net ring depth	Method (hor, obliq, vert)	Net diameter (30 or 50 cm)	Ethanol added (Y or N)	Samples combined (Y or N)	Sample sent to, date
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2) 4	Oblig			<u> </u>	
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Step 3: Collect Veliger Tows from 3 sites; the deep hole (DH) and two other deep areas along the downwind side of the lake. Submit the sample, this data form and the Mussel Veliger Tow Monitoring Report (3200-135) to DNR Science Service.

Site	Net ring depth	Net diameter (30 or 50 cm)	Ethanol added (Y or N)	Samples combined (Y or N)	Sample sent to, date	44.17463
1	24	SOCK	V	V II.		1 -44.11.800 3
2)				-84.17719
3		V	· V	<u> </u>		· ·

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