Aquatic Invasive Species (AIS) Early Detection Monitoring Data Form

Form 3200-xxx (R 5/2015)

Instructions: Bold fields must be completed

Location Name **MBIC** County S Date(s) sign? u On (ftorph)) (ZM≥99 umhos/cm) Conductivity Collector(s) **Start Time** End Time Total Hours (hrs x # ppl)

STEP 1: Circle species that you looked for and review the Identification Handout.

	Spiny/fishhook waterflea	New Zealand mudsnails	Japanese hop	Phragmites	Didymo	Parrot feather	Brazilian waterweed
	Rusty/red swamp crayfish	Asian clam	Japanese knotweed	Flowering rush	Eurasian water milfoil	Fanwort	Yellow floating heart
nails (please specify)	Chinese/Banded mystery snails	Zebra/quagga mussels	Yellow flag iris	RIPARIAN PLANTS	Water lettuce	Curly leaf pondweed	European frogbit
Other	Faucet snails	INVERTEBRATES	Purple loosestrife	Water chestnut	Water hyacinth	Hydrilla	AQUATIC PLANTS/ALGAE

sample of any new AIS found. Collect five new invasive plant specimens, 20 Dreissenids, and up to 3 of each invertebrate species. Include internal and external labels with WBIC, name of lake, county, sample date, sample type (snails, spiny water flea or zebra mussel) and collector. Legibility is appreciated. If needed, preserve with adequate

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Site*	Latitude	Longitude	Snorkel (Y/N)	Snorkel If no, indicate (Y/N) why†	Species name, density $(1-5)^{\frac{1}{5}}$, and live (L) or dead $\{D\}^{\frac{6}{5}}$	Sample (Y/N)	Photo (Y/N)	No AIS	Comments
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k0d	landing (RI) target	*host landing (RI) target site (TS), meander survey (IVIS).	survey (N	is).					

[†]Stained water, turbid water, blue-green bloom, chemical treatment, other (please describe).

^{*}Density ratings: 1-a few plants or invertebrates, 2-one or a few plant beds or colonies of invertebrates, 3-many small beds or scattered plants or colonies of invertebrates, 4-dense plant, snail, or mussel growth in a while bay or portion of the lake, or 5-dense plant, snail or mussel growth covering most shallow areas.

⁸Live (L) animals will contain flesh and live plants will generally be rooted. Dead (D) animals will not contain flesh and dead plants include sterile fragments.

completed copy of this data form, and a completed copy of the Water Flea Tow Monitoring Report (3200-128) to DNR Science Services. Legibility is appreciated. STEP 3: Collect Waterflea Tows from the deep hole (DH). Decant water and preserve the sample. Preserve with 4 parts ethanol and 1 part sample. Submit the sample, a

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		N.12/3	(Y or N)	Samples combined Date sent
C		V) 8/7/15	(Y or N)	bined
		W 8/7/15	(Y or N)	bined

STEP 4: Collect vertical Veliger Tows from 3 sites; the deep hole (DH) and two other deep areas along the downwind side of the lake. Preserve with 4 parts ethanol and 1 part sample. Submit the sample, a copy of this completed data form, and a completed copy of the Mussel Veliger Tow Monitoring Report (3200-135) to DNR Science Service. Legibility is appreciated.

Latitude .				
Longitude		-		
Net ring Net	depth (m)			
Net	depth (m) diameter†			
Ethanol*				
Samples combined Date sent	(Y or N)			-
Date sent				
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*Horizontal, oblique, or vertical

‡Non-denatured or denatured ethanol

STEP 5: Coordinate voucher and sample submission and verification with regional DNR staff for all AIS records for the specific region.

- Plants will be compiled and entered into a spreadsheet to be verified and submitted to a herbarium by an in-person appointment. Please indicate which herbarium: Freckmann Herbarium, Wisconsin State Herbarium, Other _. Date of herbarium meeting
- Snails will be compiled with other regional snail specimens and sent to UW La Crosse. Date sent
- Dreissenids will be sent to Science Services. Date sent
- Crayfish compiled and sent to: Craig Roesler or Scott VanEgeren. Date

STEP 6: Data was entered into SWIMS on β

Once data is entered, send scans of data sheets to central office (Maureen.Ferry@Wisconsin.gov and Amanda.Perdzock@Wisconsin.gov)

STEP 7: Data was proofed on

Notes: