Instructions: Bold fields must be completed.

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STEP 1: Circle species that you looked for and review the Identification Handout.

Brazilian waterweed Parrot feather Didymo Phr	Yellow floating heart Fanwort Eurasian water milfoil Flo	European frogbit Curly leaf pondweed Water lettuce RIP	AQUATIC PLANTS/ALGAE Hydrilla Water hyacinth Wa
Phragmites Japanese hop	Eurasian water milifoil Flowering rush Japanese knotweed Asian clam	RIPARIAN PLANTS Yellow flag iris	Water chestnut Purple loosestrife
 New Zealand mudsnails	Asian clam	Zebra/quagga mussels	INVERTEBRATES
sland mudsnalls Spiny/fishhook waterflea	Rusty/red swamp crayfish	Chinese/Banded mystery snails (please specify)	Faucet snails
		(please specify)	Other

WBIC, name of lake, county, sample date, sample type (snails, spiny water flea or zebra mussel) and collector. Legibility is appreciated. If needed, preserve with adequate sample of any new AIS found. Collect five new invasive plant specimens, 20 Dreissenids, and up to 3 of each invertebrate species. Include internal and external labels with STEP 2: Record locations of sampling sites (in decimal degrees). Indicate whether snorkeled or why not. List AlS found and density at each site or record none. Collect a

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^{*}boat landing (BL), target site (TS), meander survey (MS)

[†]Stained water, turbid water, blue-green bloom, chemical treatment, other (please describe).

SLive (L) animals will contain flesh and live plants will generally be rooted. Dead (D) animals will not contain flesh and dead plants include sterile fragments. *Density ratings: 1-a few plants or invertebrates, 2-one or a few plant beds or colonies of invertebrates, 3-many small beds or scattered plants or colonies of invertebrates, 4-dense plant, snail, or mussel growth in a while bay or portion of the lake, or 5-dense plant, snail or mussel growth covering most shallow areas.

STEP 3: Collect Waterflea Tows from the deep hole (DH). Decant water and preserve the sample. Preserve with 4 parts ethanol and 1 part sample. Submit the sample, a completed copy of this data form, and a completed copy of the Water Flea Tow Monitoring Report (3200-128) to DNR Science Services. Legibility is appreciated

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STEP 4: Collect vertical Veliger Tows from 3 sites; the deep hole (DH) and two other deep areas along the downwind side of the lake. Preserve with 4 parts ethanol and 1 part sample. Submit the sample, a copy of this completed data form, and a completed copy of the Mussel Veliger Tow Monitoring Report (3200-135) to DNR Science Service. Legibility is appreciated.

Latitude	Longitude	Net ring Net	Net	Ethanol [‡]	Samples combined Dat	Date sent
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^{*}Horizontal, oblique, or vertical.

130 or 50 cm.

#Non-denatured or denatured ethanol.

STEP 5: Coordinate voucher and sample submission and verification with regional DNR staff for all AIS records for the specific region.

- Freckmann Herbarium, Wisconsin State Herbarium, Other Plants will be compiled and entered into a spreadsheet to be verified and submitted to a herbarium by an in-person appointment. Please indicate which herbarium Date of herbarium meeting
- Snails will be compiled with other regional snail specimens and sent to UW La Crosse. Date sent
- Dreissenids will be sent to Science Services. Date sent ____

STEP 6: Data was entered into SWIMS on Crayfish compiled and sent to: Craig Roesler or Scott Van Egeren. Date

STEP 7: Data was proofed on Once data is entered, send scans of data sheets to central office (Maureen.Ferry@W à <u>onsin.gov</u> and <u>Amanda.Perdzock@Wisconsin.gov)</u>

Notes: