Aquatic Invasive Species (AIS) Early Detection Monitoring Data Form

Form 3200-xxx (R 5/2015)

Instructions: Bold fields must be completed.

Location Name	**WBIC**	**County**	**Date(s)**	**AIS sign?**	Secchi (ft or m)	Conductivity (ZM ≥ 99 umhos/cm)	**Collector(s)**	**Start Time**	**End Time**	**Total Hours** (hrs x # ppl)
Finnegan Lake		Oconto	6/29/15	Y	11ft	160	Michelle N. Ryan M.	11am	2pm	6

STEP 1: Circle species that you looked for and review the Identification Handout.

AQUATIC PLANTS/ALGAE	Hydrilla	Water hyacinth	Water chestnut	Purple loosestrife	**INVERTEBRATES**	Faucet snails	Other (please specify)
European frogbit	Curly leaf pondweed	Water lettuce	**RIPARIAN PLANTS**	Yellow flag iris	Zebra/quagga mussels	Chinese/Banded mystery snails	
Yellow floating heart	Fanwort	Eurasian water milfoil	Flowering rush	Japanese knotweed	Asian clam	Rusty/red swamp crayfish	
Brazilian waterweed	Parrot feather	Didymo	Phragmites	Japanese hop	New Zealand mudsnails	Spiny/fishhook waterflea	

STEP 2: Record locations of sampling sites (in decimal degrees). Indicate whether snorkeled or why not. List AIS found and density at each site or record none. Collect a sample of any new AIS found. Collect five new invasive plant specimens, 20 Dreissenids, and up to 3 of each invertebrate species. Include internal and external labels with WBIC, name of lake, county, sample date, sample type (snails, spiny water flea or zebra mussel) and collector. Legibility is appreciated. If needed, preserve with adequate ethanol.

Site*	**Latitude**	**Longitude**	**Snorkel (Y/N)**	**If no, indicate why†**	**Species name, density (1-5)‡, and live (L) or dead (D)§**	**Sample (Y/N)**	**Photo (Y/N)**	**No AIS**	**Comments**
BL	44.87810	-88.31813	Y	–	BMS - (L); EWM-2(L); CMS-1 (T. zng-1(L))	Y	N		
MS1	44.87579	-88.31943	N	–	Phrag-2(L); EWM-3(L)	Y	N		
MS2	44.87580	-88.32049	N	–	Phrag-1(L); EWM-3(L)	N	N		
MS3	44.87705	-88.31772	N	–	Phrag-1(L); EWM-1(L)	N	N		
TS1	44.87662	-88.32280	Y	–	EWM-3(L); BMS-1; CMS-1; T. zng-1 (Y) (N)				
TS2	44.87572	-88.32095	Y	–	EWM-2(L); BMS-1(L); CMS-1(L)	Y	N		
TS3	44.87740	-88.31939	Y	–	EWM-1(L); BMS-1(L); CMS-1(L)	N	N		
TS4	44.87662	-88.31825	Y	–	EWM-1(L)	N	N		
TS5	44.87578	-88.31925	Y	–	EWM-2(L); Phrag-1(L); CMS-1(L)	N	N		

*boat landing (BL), target site (TS), meander survey (MS).

†Stained water, turbid water, blue-green bloom, chemical treatment, other (please describe).

‡Density ratings: 1-a few plants or invertebrates, 2-one or a few plant beds or colonies of invertebrates, 3-many small beds or scattered plants or colonies of invertebrates, 4-dense plant, snail, or mussel growth in a while bay or portion of the lake, or 5-dense plant, snail or mussel growth covering most shallow areas.

§Live (L) animals will contain flesh and live plants will generally be rooted. Dead (D) animals will not contain flesh and dead plants include sterile fragments.

STEP 3: Collect Waterflea Tows from the deep hole (DH). Decant water and preserve the sample. Preserve with 4 parts ethanol and 1 part sample. Submit the sample, a completed copy of this data form, and a completed copy of the Water Flea Tow Monitoring Report (3200-128) to DNR Science Services. Legibility is appreciated.

Latitude	Longitude	Method*	Net ring depth (m)	Net diameter†	Ethanol‡	Samples combined (Y or N)	Date sent
44.87602	-88.32182	obl			non	Y	
44.87615	-88.32188	obl			non	Y	
44.87615	-88.32184	obl			non	Y	

STEP 4: Collect vertical Veliger Tows from 3 sites; the deep hole (DH) and two other deep areas along the downwind side of the lake. Preserve with 4 parts ethanol and 1 part sample. Submit the sample, a copy of this completed data form, and a completed copy of the Mussel Veliger Tow Monitoring Report (3200-135) to DNR Science Service. Legibility is appreciated.

Latitude	Longitude	Net ring depth (m)	Net diameter†	Ethanol‡	Samples combined (Y or N)	Date sent
44.87610	-88.32183	4		non	Y	
44.87611	-88.32164	4		non	Y	
44.87612	-88.32163	4		non	Y	

*Horizontal, oblique, or vertical.

†30 or 50 cm.

‡Non-denatured or denatured ethanol.

STEP 5: Coordinate voucher and sample submission and verification with regional DNR staff for all AIS records for the specific region.

- Plants will be compiled and entered into a spreadsheet to be verified and submitted to a herbarium by an in-person appointment. Please indicate which herbarium: Freckmann Herbarium, Wisconsin State Herbarium, Other _____. Date of herbarium meeting _____.
- Snails will be compiled with other regional snail specimens and sent to UW La Crosse. Date sent _____.
- Dreissenids will be sent to Science Services. Date sent _____.
- Crayfish compiled and sent to: Craig Roesler or Scott VanEgeren. Date _____.

STEP 6: Data was entered into SWIMS on _____ by _____

Once data is entered, send scans of data sheets to central office (Maureen.Ferry@Wisconsin.gov and Amanda.Perdzock@Wisconsin.gov).

STEP 7: Data was proofed on _____ by _____

Notes: