tower Forler

Aquatic Invasive Species (AIS) Early Detection Monitoring Data Form

Form 3200-xxx (R 5/2015)

Instructions: Bold fields must be completed

Location Name ₩BIC County Date(s) sign? AIS Secchi Conductivity
(ft or m) (ZM ≥ 99 umhos/cm) Collector(s) lenne o A. 50-Start Time **End Time** Total Hours (hrs×#.ppl)

STEP 1: Circle species that you looked for and review the Identification Handout.

European frogbit Curly leaf pondweed Water lettuce RIPARIAN PLANTS Yellow flag iris Zebra/quagga mussels Chinese/Banded mystery snails (please spe		Yellow floating heart Fanwort Eurasian water milfoil Flowering rush Japanese knotweed Asian clam Rusty/red swamp crayfish Brazilian waterweed Parrot feather Didymo Phragmites Japanese hop New Zealand mudsnails Spiny/fishhook waterflea
	(please spe	it Curly leaf pondweed Water lettuce RIPARIAN PLANTS Yellow flag iris Zebra/quagga mussels

WBIC, name of lake, county, sample date, sample type (snails, spiny water flea or zebra mussel) and collector. Legibility is appreciated. If needed, preserve with adequate sample of any new AlS found. Collect five new invasive plant specimens, 20 Dreissenids, and up to 3 of each invertebrate species. Include internal and external labels with

Site*	Site* Latitude	Longitude	Snorkel If no, (Y/N) whyt	Snorkel If no, indicate (Y/N) why†	Species name, density (1:5)*, and live
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[†]Stained water, turbid water, blue-green bloom, chemical treatment, other (please describe).

SLive (L) animals will contain flesh and live plants will generally be rooted. Dead (D) animals will not contain flesh and dead plants include sterile fragments. invertebrates, 4-dense plant, snail, or mussel growth in a while bay or portion of the lake, or 5-dense plant, snail or mussel growth covering most shallow areas. *Density ratings: 1-a few plants or invertebrates, 2-one or a few plant beds or colonies of invertebrates, 3-many small beds or scattered plants or colonies of

STEP 3: Collect Waterflea Tows from the deep hole (DH). Decant water and preserve the sample. Preserve with 4 parts ethanol and 1 part sample. Submit the sample, a completed copy of this data form, and a completed copy of the Water Flea Tow Monitoring Report (3200-128) to DNR Science Services. Legibility is appreciated

	Latitude Longitude Method* Net ring Net depth (m) diameter t	1 - 1 - 1 - 1 - 1 - 1 - 1 - 1 - 1 - 1 -
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	Samples combined (Y or N)	Vacante Ten to District
	Date sent	to play science services, regionity is appreciated
		ility is appreciated.

part sample. Submit the sample, a copy of this completed data form, and a completed copy of the Mussel Veliger Tow Monitoring Report (3200-135) to DNR Science Service. STEP 4: Collect vertical Veliger Tows from 3 sites; the deep hole (DH) and two other deep areas along the downwind side of the lake. Preserve with 4 parts ethanol and 1 Legibility is appreciated.

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depth (m) diameter	diameter†		(Y or N)	
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^{*}Horizontal, oblique, or vertical

‡Non-denatured or denatured ethanol.

STEP 5: Coordinate voucher and sample submission and verification with regional DNR staff for all AIS records for the specific region.

- Plants will be compiled and entered into a spreadsheet to be verified and submitted to a herbarium by an in-person appointment. Please indicate which herbarium: Freckmann Herbarium, Wisconsin State Herbarium, Other Date of herbarium meeting
- Snails will be compiled with other regional snail specimens and sent to UW La Crosse. Date sent
- Dreissenids will be sent to Science Services. Date sent

STEP 6: Data was entered into SWIMS on Crayfish compiled and sent to: Craig Roesler or Scott VanEgeren. Date à

Once data is entered, send scans of data sheets to central office (Maureen.Ferry@Wisconsh.gov and Amanda.Perdzock@Wisconsin.gov)

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STEP 7: Data was proofed on

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